

ACCELERATED COMMUNICATION

Aspects of Dioxin Toxicity Are Mediated by Interleukin 1-Like Cytokines

Kalyan Pande, Susan M. Moran, and Christopher A. Bradfield

McArdle Laboratory for Cancer Research, University of Wisconsin Medical School, Madison, Wisconsin

Received January 6, 2005; accepted February 18, 2005

ABSTRACT

Exposure to 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (dioxin) results in a broad spectrum of toxic effects. Most, if not all, of these responses are dependent upon the binding of dioxin to the aryl hydrocarbon receptor. Given their common roles in chemically induced toxicity, we asked whether interleukin 1 (IL1)-like cytokines play a role in acute aspects of the dioxin response. To test this idea, we employed a “triple-null” mouse model that lacks the two receptors for the tumor necrosis factors- α and - β and the receptor for the IL1- α and IL1- β

cytokines. When triple null mice were treated with dioxin, there was significant attenuation in the levels of serum alanine aminotransferase, signifying reduced hepatocellular damage. In addition, the triple-null mice were protected from dioxin-induced liver inflammation. Loss of receptors for the IL1-like cytokines was not protective for all aspects of dioxin toxicity. Endpoints such as thymic involution, *Cyp1a2* induction, hepatomegaly, and hydropic degeneration remain unchanged in this model.

The environmental pollutant 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (dioxin) causes a wide spectrum of toxic effects, including hepatocellular damage, thymic involution, teratogenesis, and cancer (Pohjanvirta and Tuomisto, 1994). The aryl hydrocarbon receptor (AHR), a member of the PAS superfamily of transcriptional regulators, mediates most, if not all, of these effects (Poland and Glover, 1980; Schmidt and Bradfield, 1996; Carver et al., 1998). The binding of dioxin to the AHR results in translocation of the receptor complex to the nucleus, where the receptor dimerizes with another basic helix-loop-helix PAS protein known as AHR nuclear translocator (ARNT) (Schmidt and Bradfield, 1996). In the nucleus, the AHR-ARNT heterodimer binds to genomic “dioxin-response elements” (DREs) and up-regulates genes encoding a battery of enzymes, including the cytochrome P450-dependent monooxygenases *Cyp1a1*, *Cyp1a2*, and *Cyp1b1* (Schmidt and Bradfield, 1996). The induction of these enzymes by the

AHR is often referred to as an adaptive response because many AHR agonists, such as benzo(*a*)pyrene, both induce this pathway and are metabolized through this response (Gu et al., 2000).

The interleukin 1 (IL1)-like cytokines are soluble factors that are known to participate in chemically-induced liver injury (Schumann and Tiegs, 1999; Yin et al., 1999). These cytokines are commonly thought of as part of the innate immunity mechanism that regulates acute phase proteins under cellular stress (Ramadori and Armbrust, 2001). The IL1-like cytokines that are released under similar physiological and pathological conditions include tumor necrosis factor (TNF) α , TNF β , IL1 α , and IL1 β (Moshage, 1997; Ramadori and Armbrust, 2001). The TNF α and TNF β cytokines can each bind to two known receptors, TNFR1 (p55) and TNFR2 (p75), encoded by the *Tnfrsf1a* and *Tnfrsf1b* loci, respectively (Ramadori and Armbrust, 2001). The IL1 α and IL1 β cytokines bind to the type I and II receptors encoded by the *Il1r1* and *Il1r2* loci, respectively. However, the binding to the type II receptor is not known to initiate signaling, and it is widely accepted that this receptor acts as a “decoy” that prevents

This work was supported by National Institutes of Health grants R01-ES012752 and P30-CA014520.

Article, publication date, and citation information can be found at <http://molpharm.aspetjournals.org>.
doi:10.1124/mol.105.010983.

ABBREVIATIONS: Dioxin, 2,3,7,8-tetrachlorodibenzo-*p*-dioxin; AHR, aryl hydrocarbon receptor; PAS, period-ARNT-single-minded; ARNT, aryl hydrocarbon receptor nuclear translocator; DRE, dioxin-response element; IL, interleukin; TNF, tumor necrosis factor; TNFR, tumor necrosis factor receptor; ALT, alanine aminotransferase; PCR, polymerase chain reaction; DMSO, dimethyl sulfoxide; TUNEL, terminal deoxynucleotidyl transferase dUTP nick-end labeling; H&E, hematoxylin and eosin.

IL1 binding to the type I receptor (Glaccum et al., 1997). The IL1-like cytokines act in both autocrine and paracrine manners by binding to their respective receptors on target cells, resulting in the initiation of various intracellular signaling cascades. An example of this is the up-regulation of the nuclear factor κ b and the mitogen-activated protein kinase pathways that are induced in acute phase response by both TNF and IL1 (Moshage, 1997; Ramadori and Armbrust, 2001).

The common involvement of IL1-like cytokines in forms of chemically induced liver injury led us to hypothesize that these cytokines are important downstream mediators of certain aspects of dioxin-induced hepatotoxicity. For example, TNF α has been shown to participate in the acute liver injury that occurs in response to concanavalin A, endotoxin, and alcohol (Bradham et al., 1998; McClain et al., 1998; Yin et al., 1999; Diehl, 2000). Both TNF α and IL1 α have also been shown to play a role in acetaminophen-induced liver toxicity (Blazka et al., 1996). In consideration of the overlapping patterns of intracellular signaling and gene expression activated by the various IL1 and TNF agonists and receptors, we generated a "triple-null" mouse line that is deficient for all known IL1 and TNF signaling. This was accomplished by generating compound mutants lacking all three functional receptors (i.e., TNFR1, TNFR2, and IL1R1). In this study, we used this triple-null mouse model to identify those aspects of dioxin-induced toxicity that are dependent on IL1-like cytokine signaling.

Materials and Methods

Animals. Animals were housed in a specific pathogen-free facility on corn cob bedding with food and water ad libitum following the rules and regulations established by the University of Wisconsin Animal Care and Use Committee.

Generation of "Triple-Null" Mice Lacking *Tnfrsf1a*, *Tnfrsf1b*, and *Il1r1*. We obtained mice that harbor null alleles at the *Tnfrsf1a*, *Tnfrsf1b*, and *Il1r1* loci from the Jackson Laboratory (Bar Harbor, ME) (Pfeffer et al., 1993; Erickson et al., 1994; Glaccum et al., 1997). Although we use the abbreviated nomenclature above, the specific nomenclature for the *Tnfrsf1a*, *Tnfrsf1b*, and *Il1r1* lines is B6.129-*Tnfrsf1a*^{tm1Mak}/J, B6.129S2-*Tnfrsf1b*^{tm1Mwm}/J, and B6.129S7-*Il1r1*^{tm1Imx}/J, respectively (<http://www.informatics.jax.org/mgihome/nomen/strains.shtml>). To generate the triple-null mouse, *Tnfrsf1a* $-/-$ and *Tnfrsf1b* $-/-$ mice were first interbred to generate a *Tnfrsf1a* $-/-$ /*Tnfrsf1b* $-/-$ double-null line. This line was then crossed with an *Il1r1* $-/-$ mouse to generate a *Tnfrsf1a* $-/-$ /*Tnfrsf1b* $-/-$ /*Il1r1* $-/-$ triple-null mouse line. Mice were genotyped by PCR protocols established for the individual alleles using genomic DNA isolated from tail biopsies (Pfeffer et al., 1993; Erickson et al., 1994; Glaccum et al., 1997). In addition, RT-PCR was carried out to examine expression from the individual alleles using liver total RNA (RNeasy Midi Kit; QIAGEN, Valencia, CA). The primer sets used for the RT-PCR were: *Tnfrsf1a* allele, forward primer OL 5655 (5'-GCTGTGTCCCAAGGAAAG) and reverse primer OL 5656 (5'-GTCCTGGGGTTTGTGACAT); *Tnfrsf1b* allele, forward primer OL 5659 (5'-TCTAGCTCCAGGCACAAGG) and reverse primer OL 5660 (5'-CATTTGGGGCTCTTGAAGT); *Il1r1* allele, forward primer OL 5662 (5'-TTGAGGAGGCAGTTTCGTT) and reverse primer OL 5663 (5'-TACCACTGGACCTCGGGTAA). Because individual receptor null animals had been crossed to the C57BL/6J (B6) background for at least five generations before import, the triple-null line used in these studies is more than 95% B6 background.

Dioxin Hepatotoxicity. At 7 to 8 weeks of age, male mice were intraperitoneally injected with either 64 μ g/kg dioxin or the same

volume of vehicle alone (DMSO). Fourteen days after the dioxin treatment, mice were anesthetized using isoflurane, and blood was obtained by cardiac puncture. Liver and thymus weights were then measured and normalized to whole body weights. Medial sections from the left lobe of the livers were fixed in 10% (w/v) buffered formalin for histological examination. Apoptosis was determined by terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling (TUNEL) assay using the in situ cell death detection kit POD (Roche Applied Science, Indianapolis, IN). The number of TUNEL-positive cells was estimated using Stereo Investigator software (MicroBrightfield Inc., Williston, VT). To visualize macrophage/monocyte populations, immunostaining was performed with rat anti-mouse F4/80 monoclonal antibody (Serotec Inc., Raleigh, NC), biotinylated rabbit anti-rat secondary antibody (Vector Labs, Burlingame, CA), and avidin-biotin conjugated system with NovaRED substrate (Vector Labs, Burlingame, CA) (Martinez-Pomares et al., 1996). The number of inflammatory macrophages was estimated from F4/80 immunostained slides using Stereo Investigator software (MicroBrightfield Inc.). Sections stained with hematoxylin and eosin (H&E) were used to estimate the percentage of cells displaying hydropic degeneration as follows: 0 (absent); 1 (rare, scattered); 2 (below 25%); 3 (25–50%); 4 (50–75%); 5 (75–100%). Alanine aminotransferase (ALT) activity was determined on sera by the Clinical Pathology Laboratory at the University of Wisconsin, School of Veterinary Medicine in Madison. Statistical analysis was performed using an ANOVA, and Tukey's test was used to determine differences with $p < 0.05$.

Northern Analysis. *Cyp1a2* expression was determined with Northern analysis using 10 μ g of total RNA. The CYP1A2 probe (Plasmid 1232) was labeled with [³²P]dCTP using Rediprime II Random Primer DNA Labeling System (Amersham Biosciences, UK).

Results

Generation of Mice Deficient for *Tnfrsf1a*, *Tnfrsf1b*, and *Il1r1*. To determine whether IL1-like cytokines are important in dioxin toxicity, we generated mice lacking the three receptors for the four IL1-like cytokines—IL1 α , IL1 β , TNF α , and TNF β . The triple-null mouse was generated by intercrossing mice having individual null alleles until animals homozygous at all three null loci were obtained. To confirm the presence of all three null alleles, a PCR genotyping approach was performed based on published protocols (Pfeffer et al., 1993; Erickson et al., 1994; Glaccum et al., 1997). To demonstrate that each targeted allele resulted in disruption in corresponding gene expression, RT-PCR was performed on liver mRNA from the triple-null and wild-type mice (Fig. 1). For *Tnfrsf1a* and *Tnfrsf1b*, we designed primers that amplify the mRNA corresponding to exons 2 to 6 and

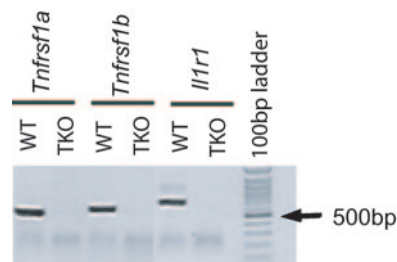


Fig. 1. All three IL1-like receptors are absent in triple-null mice. The RT-PCR was performed for each allele to confirm the loss of all three receptors in the triple-null. Lanes 1 and 2 show *Tnfrsf1a* mRNA amplification in wild-type and triple-null mice, respectively. Lanes 3 and 4 show *Tnfrsf1b* mRNA amplification in wild-type and triple-null mice, respectively. Lanes 5 and 6 show *Il1r1* mRNA amplification in wild-type and triple-null mice, respectively.

exons 1 to 5, respectively. For *Il1r1*, the mRNA region corresponding to exons 1 to 4 was amplified. The RT-PCR products for all three loci were positive for the expected sizes in wild-type mice but not in triple-null mice (Fig. 1). Although mating efficiency is reduced in triple-null mice, the adult animals display body weights and liver histology that is similar to those of their wild-type control mice (data not shown). Given that most null alleles are derived from 129-derived ES cells, we genotyped the triple null strain at the *Ahr* locus and confirmed that they possess the high-affinity allele *Ahr^{b-1}* commonly found in B6 strain, and not the low-affinity allele *Ahr^d*, commonly found in 129 strain (data not shown) (Poland et al., 1987).

Hepatomegaly, Thymic Involution, Hepatic Hydroptic Changes, and *Cyp1a2* Induction Are Unchanged in Dioxin-Exposed Triple-Null Mice. Both wild-type and triple-null mice exposed to dioxin at 64 $\mu\text{g}/\text{kg}$ display similar levels of hepatomegaly, thymic involution, and hepatic hydroptic changes (Fig. 2, A–C). The relative liver weights in wild-type mice increased by 30%, whereas the triple-null relative liver weight increased 32% in response to dioxin (Fig. 2A). Likewise, the wild-type thymus weight decreased 72% and the thymus weight in triple-null mice was reduced 69% as a result of dioxin exposure (Fig. 2B). Dioxin exposure also caused similar levels of hydroptic degeneration in both wild-type and triple-null mice at 14 days (Fig. 2C). We define hydroptic degeneration as vacuoles in the cytoplasm of hepatocytes that meet three criteria. First, they have a ragged perimeter. Second, they are negative for lipid by oil-red-o staining. Third, they are negative for glycogen as determined by PAS staining. A qualitative scoring of H&E sections from all liver sections did not reveal even a subtle difference when comparing control wild-type and triple-null mice or when comparing dioxin-treated wild-type and triple-null mice (Fig. 2, C and D). Induction of the *Cyp1a2* gene

product was used as a classic response to dioxin-activated AHR signaling. We assessed *Cyp1a2* expression using Northern analysis in each of the four treatment groups (Fig. 2E). In both wild-type and triple-null mice, dioxin treatment resulted in similar levels of induction of the *Cyp1a2* gene.

Triple Null Mice Are Protected from Dioxin-Induced Hepatocellular Damage. We observed that dioxin exposure led to infiltration of inflammatory cells in the livers of wild-type mice (Fig. 3, A and B). The types of inflammatory cells identified include macrophages, monocytes, and smaller numbers of lymphocytes and neutrophils. The density of infiltrating F4/80 positive macrophages and monocytes within the liver increased by more than 23-fold after 64 $\mu\text{g}/\text{kg}$ dioxin treatment of wild-type mice. When comparing wild-type mice, this difference between dioxin-treated and control was significantly different at $p < 0.001$ (Fig. 3B). In triple-null mice, dioxin-induced increase in the density of macrophages and monocytes was also observed. Yet this increase was not statistically different from the corresponding control group at $p < 0.05$ (Fig. 3B). It is noteworthy that dioxin-treated wild-type mice exhibited approximately 4-fold greater density of infiltrating macrophages compared with dioxin-treated triple-null mice. This difference was found to be statistically significant at $p < 0.01$.

We also observed that dioxin causes significant hepatocellular damage in wild-type mice, as indicated by an elevation of more than 21-fold in serum ALT levels (Fig. 3C). In contrast, the elevation of serum ALT in triple-null mice exposed to dioxin was only ~4-fold of control, and the difference was not statistically significant at $p < 0.05$ (Fig. 3C).

Dioxin-Induced Hepatocyte Apoptosis Is Attenuated in Triple-Null Mice. In dioxin-exposed wild-type mice, we observed induction of hepatocyte apoptosis in regions corresponding to regions of inflammatory cell infiltration (Fig. 4A). Therefore, we used TUNEL staining as an additional

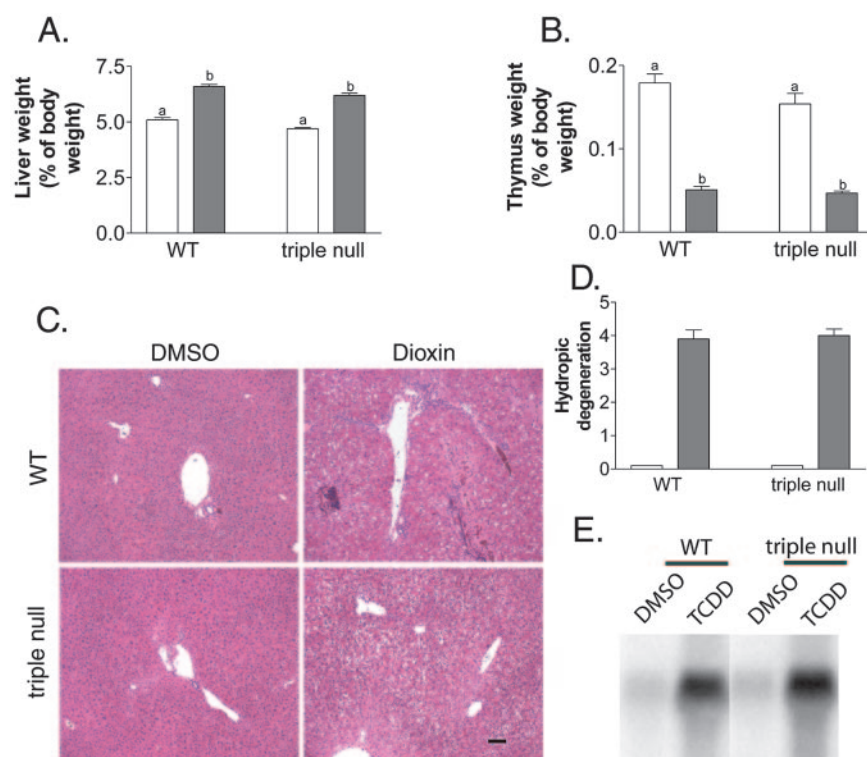


Fig. 2. Dioxin induces hepatomegaly, thymic involution, and *Cyp1a2* induction in both wild-type and triple null mice. Male mice (6–8 weeks old) were injected intraperitoneally with either 64 $\mu\text{g}/\text{kg}$ dioxin dissolved in DMSO (vehicle) or an equal volume of DMSO alone. Fourteen days after dioxin treatment, liver and thymus weights were measured and normalized to whole body weights (A and B). Medial sections from the left lobe of the liver were fixed in 10% formalin, and sections were prepared and stained with H&E to assess the overall liver damage (C). The H&E-stained slides were then scored for hydroptic degeneration (D). Total RNA from the liver was used to perform Northern analysis to detect the *Cyp1a2* mRNA (E). Scale, 100 μm . White bars, vehicle-treated animals; gray bars, dioxin-treated animals. Error bars, S.E.M. Those groups not sharing a superscript letter differ significantly at $p < 0.001$. The number of animals in each group is at least four.

measure of dioxin-induced hepatocellular toxicity and an indirect measure of liver inflammation. Dioxin caused a 10-fold increase in hepatocyte apoptosis in wild-type mice, and this difference was significant at $p < 0.001$ (Fig. 4, B and C). Likewise, the difference between dioxin-treated wild-type mice and dioxin-treated triple-null mice was significant at $p < 0.001$. In contrast, only marginal apoptosis was seen in triple-null mice treated with dioxin, and the difference between control and dioxin-treated mice was not significant at $p < 0.05$ (Fig. 4C).

Discussion

To demonstrate a role of IL1-like cytokines in dioxin toxicology, we generated a triple-null mouse line that is deficient in the receptors for $\text{TNF}\alpha$, $\text{TNF}\beta$, $\text{IL1}\alpha$, and $\text{IL1}\beta$. Exposure of these mice to dioxin demonstrated that IL1-like cytokines are involved in particular aspects of dioxin toxicity. We observed that dioxin-induced hepatocellular damage, as assessed by serum ALT levels, hepatic infiltration by inflammatory cells, and hepatocyte apoptosis were all significantly reduced in triple-null animals. Despite the significant pro-

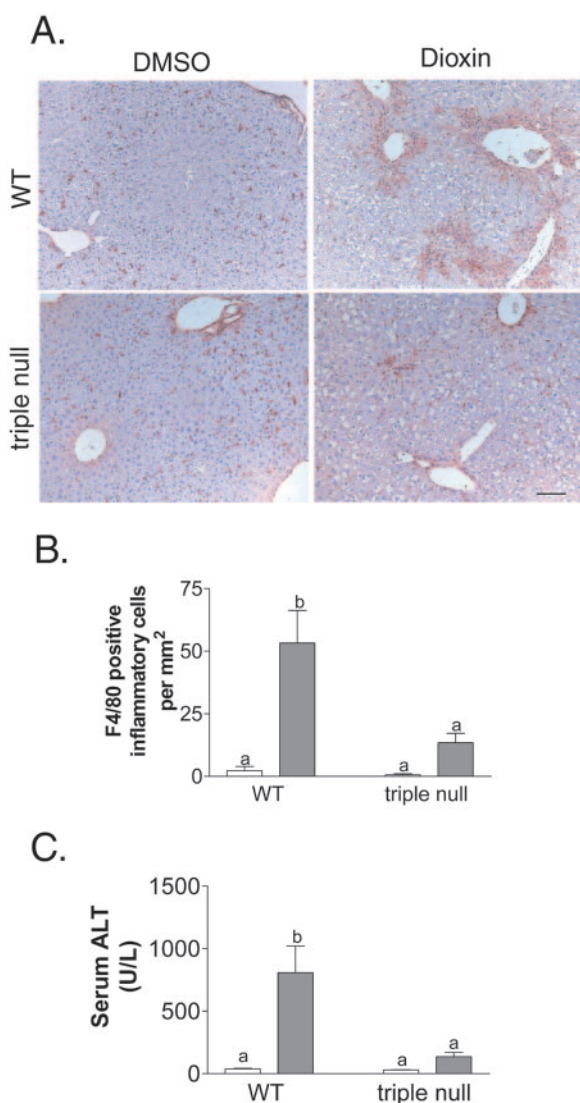


Fig. 3. Triple-null mice are protected from dioxin-induced hepatocellular damage. Male mice (6–8 weeks old) were injected intraperitoneally with either 64 $\mu\text{g/kg}$ dioxin dissolved in DMSO or an equal volume of DMSO alone. After 14 days, medial sections from the left lobe of the liver were fixed in 10% formalin, and the sections were prepared and stained with F4/80 antibody to count the number of macrophages and monocytes within inflammatory cell clusters (A). The extent of dioxin-induced inflammatory cell infiltration in the triple-null mice is reduced compared with that in wild-type mice (B). Blood sera were also obtained from mice for measurement of ALT levels (C). In wild-type mice, dioxin causes significant ALT induction, whereas this induction is not significant in triple-null mice. Scale, 100 μm . White bars, vehicle-treated animals; gray bars, dioxin-treated animals. Error bars, S.E.M. Those groups not sharing a superscript letter differ significantly at $p < 0.01$. The number of animals in each group is at least four.

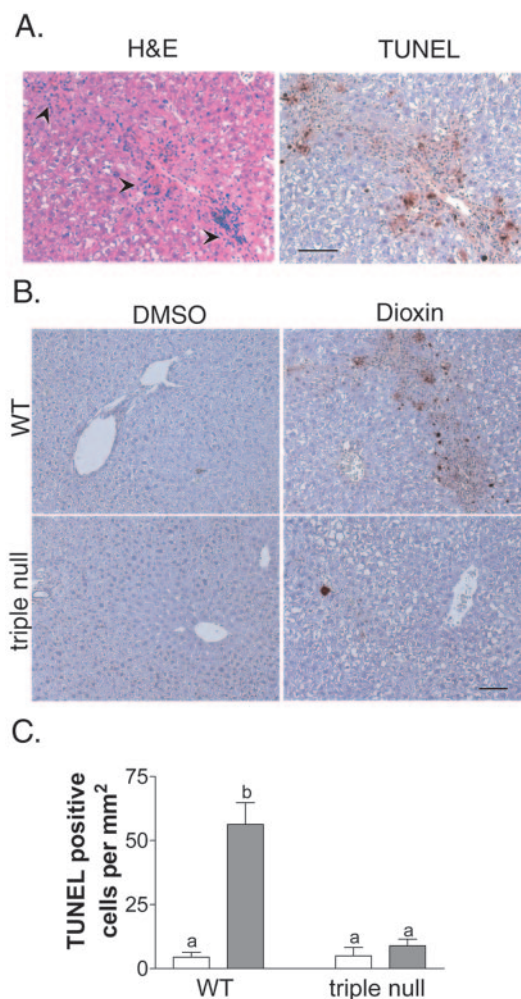


Fig. 4. Dioxin-induced hepatocyte apoptosis is reduced in triple null mice. Male mice (6–8 weeks old) were injected intraperitoneally with either 64 $\mu\text{g/kg}$ dioxin dissolved in DMSO (vehicle) or an equal volume of DMSO alone. After 14 days, medial sections from the left lobe of the dioxin-treated livers from wild-type mice were fixed in 10% formalin, and the sections were prepared and stained with H&E (A). The TUNEL assay was performed on adjacent liver sections for assessment of hepatocyte apoptosis. Comparison between H&E- and TUNEL-stained sections in dioxin-treated livers from wild-type mice reveal a high degree of correlation between the sites of inflammatory cells (arrowheads) and hepatocyte apoptosis (A). TUNEL staining was also performed on control and dioxin-treated livers from both wild-type and triple-null mice (B). Significant hepatocyte apoptosis is seen after dioxin treatment in wild-type mice but not in triple null-mice. The TUNEL-positive cells were estimated using the Stereo Investigator software. In control mice, dioxin induces significant apoptosis, whereas this induction is absent in triple null mice (C). Scale, 100 μm . White bars, vehicle treated animals; gray bars, dioxin treated animals. Error bars, S.E.M. Those groups not sharing a superscript letter differ significantly at $p < 0.001$. The number of animals in each group is at least three.

tection for these endpoints, the triple-null genotype had no influence on dioxin-induced thymic involution, hepatomegaly, *Cyp1a2* induction, or hydropic degeneration. Taken in sum, these data support the idea that some but not all responses to dioxin are dependent upon signaling by the IL1-like cytokines.

These observations are in keeping with the idea that dioxin toxicity is multifactorial and that distinct sets of signaling events lead to different toxic endpoints. One example of this idea is the observation that dioxin-induced hepatomegaly occurs normally in triple-null mice, whereas other toxic endpoints are reduced. One model to explain this result is that hepatomegaly is a direct consequence of the dioxin-induced P450 enzymes. The coordinated proliferation of smooth endoplasmic reticulum may underlie increased hepatocyte size and concomitant hepatomegaly (Schulte-Hermann, 1974; Turner and Collins, 1983). These data also support a model in which dioxin hepatotoxicity is a progressive event, where P450 induction, hepatomegaly and hydropic degeneration represent the earlier stages of acute exposure, whereas hepatocellular damage and inflammatory cell infiltration are sequelae that occur at a later stage. In such a model, hydropic degeneration might represent a crucial state in which hepatocyte physiology is significantly disrupted. We propose that this disruption results in IL1-like cytokine release by resident macrophages, subsequent inflammation, and further hepatocellular damage. It follows that removal of the IL1-like signal would halt this progression. Thus dioxin exposure would result in P450 induction, hepatomegaly, and hydropic degeneration, but not hepatocellular damage and inflammatory cell infiltration.

The IL1-like cytokines, $TNF\alpha$, $TNF\beta$, $IL1\alpha$, and $IL1\beta$, are soluble factors that are known to participate in various aspects of liver injury. The classification of cytokines as "IL1-like" comes from observations of the "acute phase response," where signaling by the IL1-like cytokines can be distinguished from responses stimulated by the IL6-like cytokines. The signaling of IL1-like cytokines results in the induction of type I acute phase proteins (Ramadori and Armbrust, 2001). The three IL1-like cytokine receptors, $TNFR1$, $TNFR2$, and $IL1R1$, are able to activate both overlapping and distinct intracellular pathways (Moshage, 1997; Ramadori and Armbrust, 2001). Because of this overlap in signaling, we chose to develop an animal model that was null for all three receptors and therefore incapable of mediating any IL1-like response. The primary advantage of this model is its greater sensitivity in detecting IL1-like signaling, because pathway redundancy has been reduced through the generation of a compound null.

Although we have not found this to be reported previously, we observed that the regions of dioxin-induced hepatocyte apoptosis correlated with sites of inflammatory cells in treated livers. This overlap provides evidence that inflammation and apoptosis are related; however, it is unclear which event precedes the other. It is possible that hepatocyte apoptosis is not as clean as previously thought and that it triggers infiltration of inflammatory cells that provide support in clearing up the debris resulting from cell death (Maher and Gores, 1998). On the other hand, the recruitment of inflammatory cells to the liver could lead to hepatocellular stress, leading to hepatocellular apoptosis. It is also possible that the two effects amplify each other (i.e., the presence of inflammatory cells result in cellular death; this, in turn, re-

cruits more inflammatory cells to that site). Further experiments are needed to explain which, if any, of these scenarios occur in dioxin-treated animals.

The rationale for our hypothesis that IL1-like cytokines may be involved in dioxin toxicity came from a broad spectrum of experiments that have been published in the last decade. Foremost is the observation that IL1-like cytokines have been shown to be common mediators of the hepatotoxicity induced by a number of agents, including alcohol, acetaminophen, endotoxin, and concanavalin A (Blazka et al., 1996; Bradham et al., 1998; McClain et al., 1998; Yin et al., 1999; Diehl, 2000). Second, a number of inflammatory cytokines have been linked to dioxin toxicity by other laboratories (Taylor et al., 1992; Moos et al., 1994; Charles and Shiverick, 1997; Fan et al., 1997; Fisher et al., 2004). In this regard, dioxin has been shown to induce $TNF\alpha$ and $IL1\beta$ mRNAs, supporting the possibility that the expression of these cytokines are DRE-mediated (Fan et al., 1997). Although direct AHR regulation is a formal possibility, our observations are also consistent with a model in which these cytokines respond to dioxin indirectly. One model compatible with our results is that P450 induction, hepatomegaly and hydropic degeneration represent primary effects of dioxin. These events could subsequently lead to the disrupted physiology of hepatocytes and the activation of IL1-like cytokines. The cytokine action could then result in the secondary effects, namely hepatocyte damage and infiltration of inflammatory cells.

Using a triple-null mouse model in which TNF and $IL1$ signaling are abrogated, we demonstrated clear roles for IL1-like cytokines in mediating aspects of dioxin-induced hepatotoxicity. In addition to its importance in dioxin toxicity, we expect this mouse model to be very useful in other areas of toxicology. In future studies, we plan to address two important questions. First, which subsets of the IL1-like cytokine pathways are essential for these toxic responses (e.g., is it $TNF\alpha$ alone?) and, second, what other aspects of dioxin toxicity are mediated by IL1-like cytokines (e.g., tumor promotion).

Acknowledgments

We thank Dr. Henry Pitot for expert histological analysis and Bernice Lin and Linh Nguyen for help with tissue collection, and Ed Glover for injections.

References

- Blazka ME, Elwell MR, Holladay SD, Wilson RE, and Luster MI (1996) Histopathology of acetaminophen-induced liver changes: role of interleukin 1 alpha and tumor necrosis factor alpha. *Toxicol Pathol* **24**:181-189.
- Bradham CA, Plumpe J, Manns MP, Brenner DA, and Trautwein C (1998) Mechanisms of hepatic toxicity. I. TNF -induced liver injury. *Am J Physiol* **275**:G387-G392.
- Carver LA, LaPres JJ, Jain S, Dunham EE, and Bradfield CA (1998) Characterization of the Ah Receptor-associated protein, ARA9. *J Biol Chem* **273**:33580-33589.
- Charles GD and Shiverick KT (1997) 2,3,7,8-Tetrachlorodibenzo-p-dioxin increases mRNA levels for interleukin-1beta, urokinase plasminogen activator and tumor necrosis factor-alpha in human uterine endometrial adenocarcinoma RL95-2 cells. *Biochem Biophys Res Commun* **238**:338-342.
- Diehl AM (2000) Cytokine regulation of liver injury and repair. *Immunological Rev* **174**:160-171.
- Erickson SL, de Sauvage FJ, Kikly K, Carver-Moore K, Pitts-Meek S, Gillett N, Sheehan KC, Schreiber RD, Goeddel DV, and Moore MW (1994) Decreased sensitivity to tumour-necrosis factor but normal T-cell development in TNF receptor-2-deficient mice. *Nature (Lond)* **372**:560-563.
- Fan F, Yan B, Wood G, Viluksela M, and Rozman KK (1997) Cytokines ($IL-1\beta$ and $TNF\alpha$) in relation to biochemical and immunological effects of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) in rats. *Toxicology* **116**:9-16.
- Fisher MT, Nagarkatti M, and Nagarkatti PS (2004) Combined screening of thymo-

- cytes using apoptosis-specific cDNA array and promoter analysis yields novel gene targets mediating TCDD-induced toxicity. *Toxicol Sci* **78**:116–124.
- Glaccum MB, Stocking KL, Charrier K, Smith JL, Willis CR, Maliszewski C, Livingston DJ, Peschon JJ, and Morrissey PJ (1997) Phenotypic and functional characterization of mice that lack the type I receptor for IL-1. *J Immunol* **159**: 3364–3371.
- Gu Y-Z, Hogenesch JB, and Bradfield CA (2000) The PAS superfamily: sensors of environmental and developmental signals. *Annu Rev Pharmacol Toxicol* **40**:519–561.
- Maier JJ and Gores GJ (1998) Apoptosis: silent killer or neutron bomb? *Hepatology* **28**:865–867.
- Martinez-Pomares L, Platt N, McKnight AJ, da Silva RP, and Gordon S (1996) Macrophage membrane molecules: markers of tissue differentiation and heterogeneity. *Immunobiology* **195**:407–416.
- McClain CJ, Barve S, Barve S, Deaciuc I, and Hill DB (1998) Tumor necrosis factor and alcoholic liver disease. *Alcohol Clin Exp Res* **22**:248S–252S.
- Moos AB, Baecher-Steppan L, and Kerkvliet NI (1994) Acute inflammatory response to sheep red blood cells in mice treated with 2,3,7,8-tetrachlorodibenzo-p-dioxin: the role of proinflammatory cytokines, IL-1 and TNF. *Toxicol Appl Pharmacol* **127**:331–335.
- Moshage H (1997) Cytokines and the hepatic acute phase response. *J Pathol* **181**: 257–266.
- Pfeffer K, Matsuyama T, Kundig TM, Wakeham A, Kishihara K, Shahinian A, Wiegmann K, Ohashi PS, Kronke M, and Mak TW (1993) Mice deficient for the 55 kd tumor necrosis factor receptor are resistant to endotoxic shock, yet succumb to L. monocytogenes infection. *Cell* **73**:457–467.
- Pohjanvirta R and Tuomisto J (1994) Short-term toxicity of 2,3,7,8-tetrachlorodibenzo-p-dioxin in laboratory animals: effects, mechanisms and animal models. *Pharmacol Rev* **46**:483–549.
- Poland A and Glover E (1980) 2,3,7,8-Tetrachlorodibenzo-p-dioxin: segregation of toxicity with the Ah locus. *Mol Pharmacol* **17**:86–94.
- Poland A, Glover E, and Taylor BA (1987) The murine Ah locus: a new allele and mapping to chromosome 12. *Mol Pharmacol* **32**:471–478.
- Ramadori G and Armbrust T (2001) Cytokines in the liver. *Eur J Gastroenterol Hepatol* **13**:777–784.
- Schmidt JV and Bradfield CA (1996) Ah receptor signaling pathways. *Annu Rev Cell Dev Biol* **12**:55–89.
- Schulte-Hermann R (1974) Induction of liver growth by xenobiotic compounds and other stimuli. *CRC Crit Rev Toxicol* **3**:97–158.
- Schumann J and Tiegs G (1999) Pathophysiological mechanisms of TNF during intoxication with natural or man-made toxins. *Toxicology* **138**:103–126.
- Taylor MJ, Lucier GW, Mahler JF, Thompson M, Lockhart AC, and Clark GC (1992) Inhibition of acute TCDD toxicity by treatment with anti-tumor necrosis factor antibody or dexamethasone. *Toxicol Appl Pharmacol* **117**:126–132.
- Turner JN and Collins DN (1983) Liver morphology in guinea pigs administered either pyrolysis products of a polychlorinated biphenyl transformer fluid or 2,3,7,8-tetrachlorodibenzo-p-dioxin. *Toxicol Appl Pharmacol* **67**:417–429.
- Yin M, Wheeler MD, Kono H, Bradford BU, Gallucci RM, Luster MI, and Thurman RG (1999) Essential role of tumor necrosis factor alpha in alcohol-induced liver injury in mice. *Gastroenterology* **117**:942–952.

Address correspondence to: Christopher A. Bradfield, McArdle Laboratory for Cancer Research, 1400 University Avenue, Madison, WI 53706-1599. E-mail: bradfield@oncology.wisc.edu
